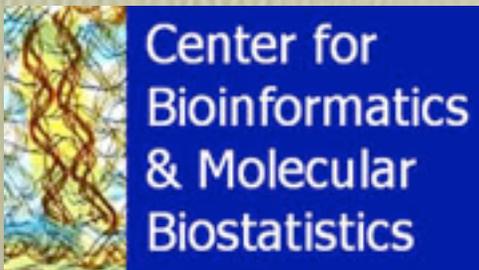


# A Principal Curve Approach to Three-Dimensional Chromatin Configuration Reconstruction

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BIRS 2018, Oaxaca

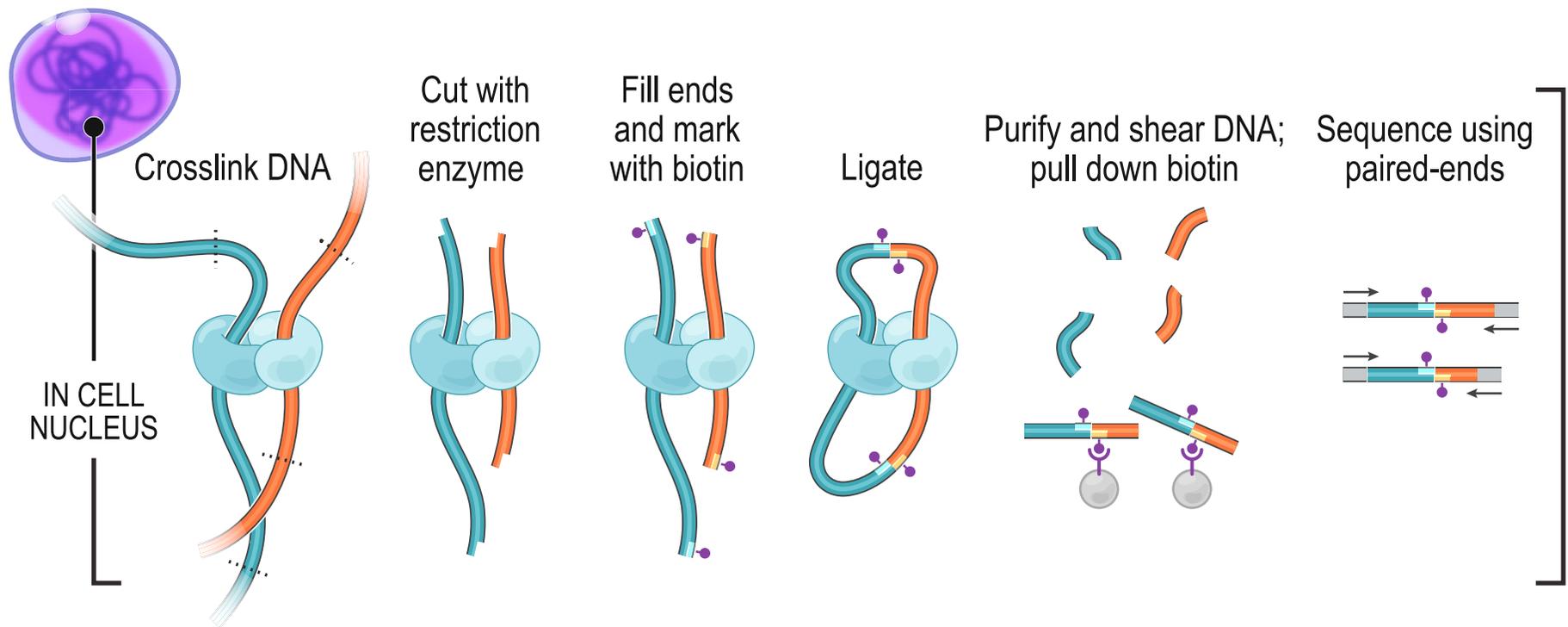
# Importance of 3D Conformation

- Gene regulation:
  - **co-localization** of co-expressed genes into transcription factories
  - **positioning** of distal control elements
- Translocations / gene fusions:
  - 20% of human cancer morbidity
  - 3D structure “**probably pivotal**”

# Observing / Inferring 3D Structure

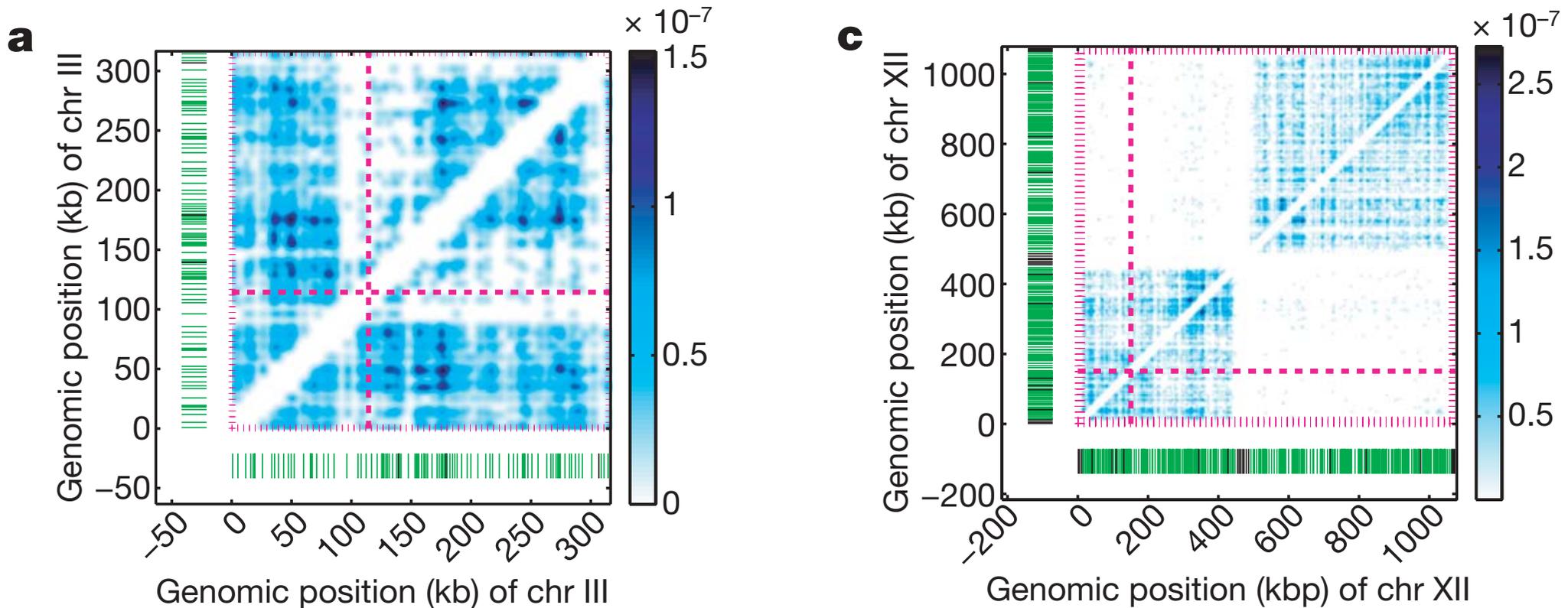
- Challenging at even modest resolutions:
  - genomes are highly condensed
  - genomes are dynamic, variable
  - traditional assays are low throughput and low resolution (**FISH coarse**)
- Recently devised suite of **C**hromatin **C**onformation **C**apture techniques has

# 3C / 4C / 5C / Hi-C / TCC



Performed using large –  $\sim 10^6$  – cell populations

# Output: Contact / Interaction Maps



Also *inter*-chromosomal maps.

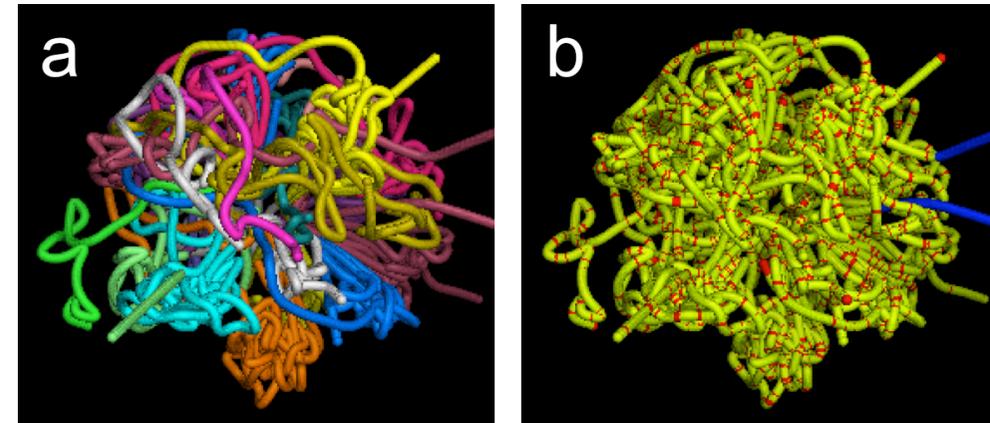
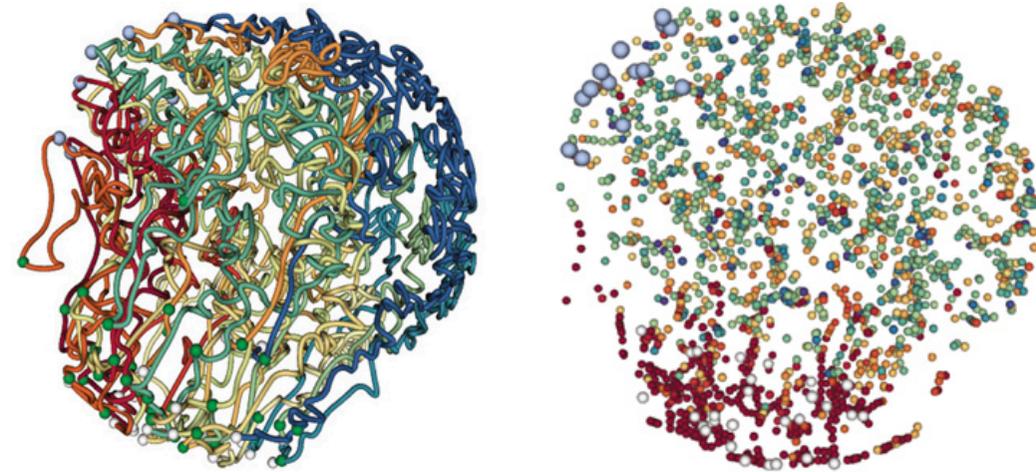
Resolution determined by binning.

# From Contacts to 3D Structure

- Objective: given contact matrix  $C$ , obtain a 3D structure (or an ensemble thereof) the between-loci pairwise distances of which recapitulate corresponding contact counts.
- Many reconstruction algorithms advanced.
- Despite assumptions, uncertainties added value derives from inferring 3D architecture:
  - In part derives from super-posing genome attributes on the reconstruction.

# 3D *P. falciparum* : Overlaid Expression

# 3D *S. cerevisiae* : Overlaid ChIP-Seq

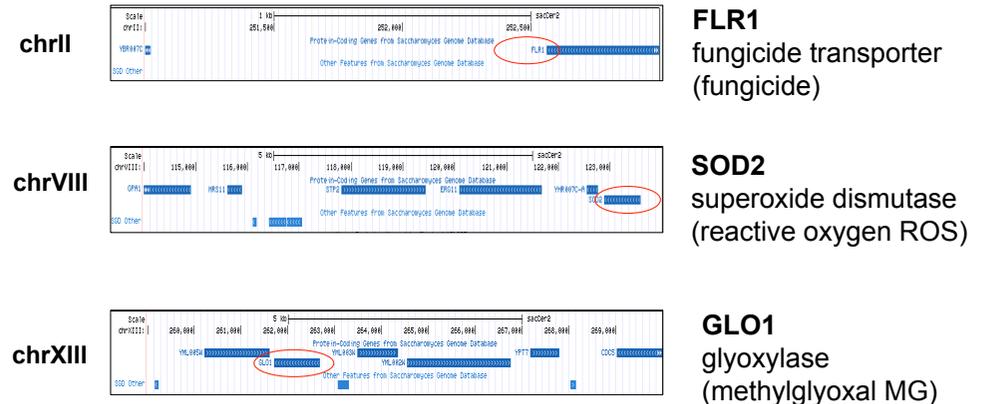


**swi6\_minbeads25\_box18**

3 regions from 3 chromosomes

chrII:	251 kB – 252 kB	(3 beads)
chrVIII:	114 kB - 124 kB	(21 beads)
chrXIII:	259 kB – 270 kB	(21 beads)

Can demonstrate expression-  
telomere distance gradient.  
But what about detecting  
focal regions : **3D hotspots** ?



# Optimization / Consensus Methods

- Generally utilize two steps:
  - convert  $C$  into a distance matrix  $D$  that captures expected pairwise distances
    - differing assumptions;  $D \propto C^{-\alpha}$ ;  $\alpha > 0$
    - sometimes interplay with second step
  - learn / estimate 3D structure from  $D$ 
    - multi-dimensional scaling (MDS) criteria
    - weights, non-metric, **constraints**, ...
    - algorithms include SA, IPO, SDE, MM...

# Distances to 3D Structure

- Minimize objective function that places (as much as possible) interacting loci at their expected distance apart (**MDS**):

$$\min_{\{x_i, x_j \in R^3\}} \sum_{\{i, j | D_{ij} < \infty\}} \omega_{ij} \cdot (\|x_i - x_j\| - D_{ij})^2$$

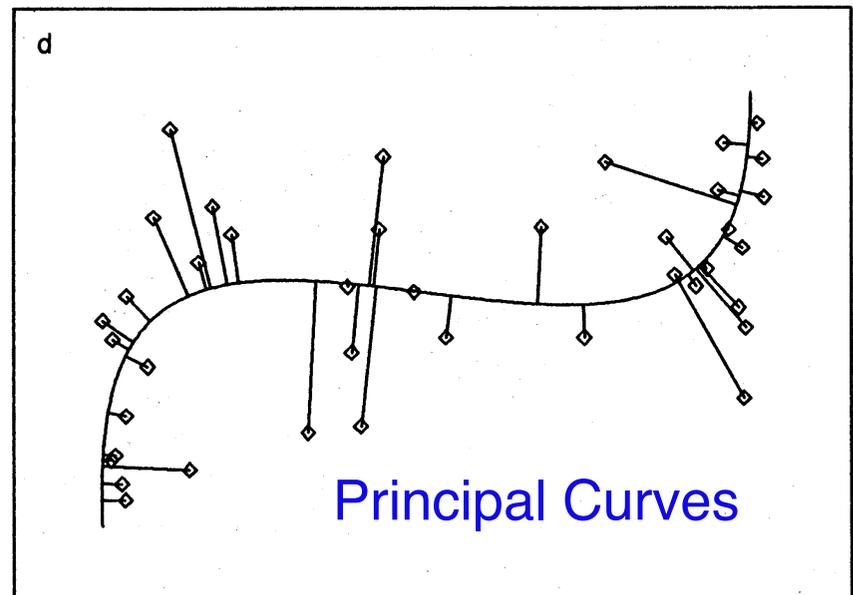
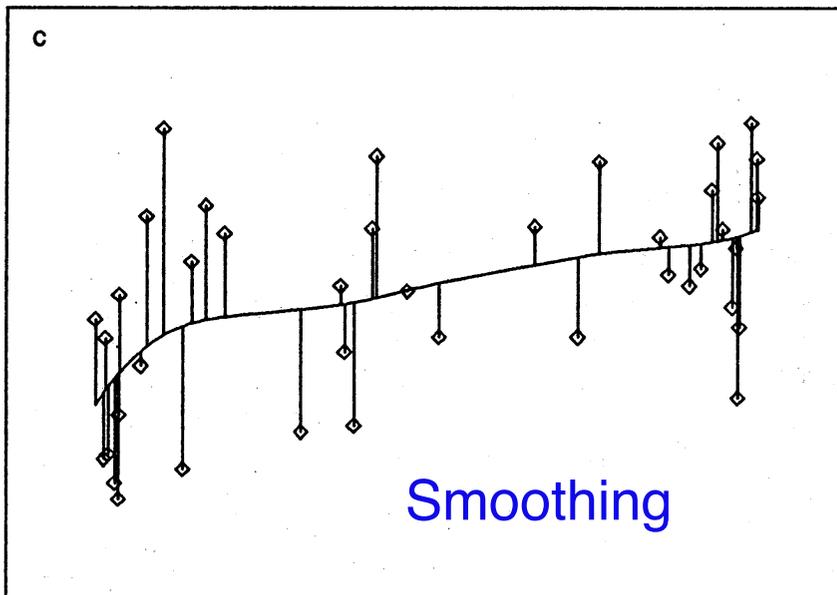
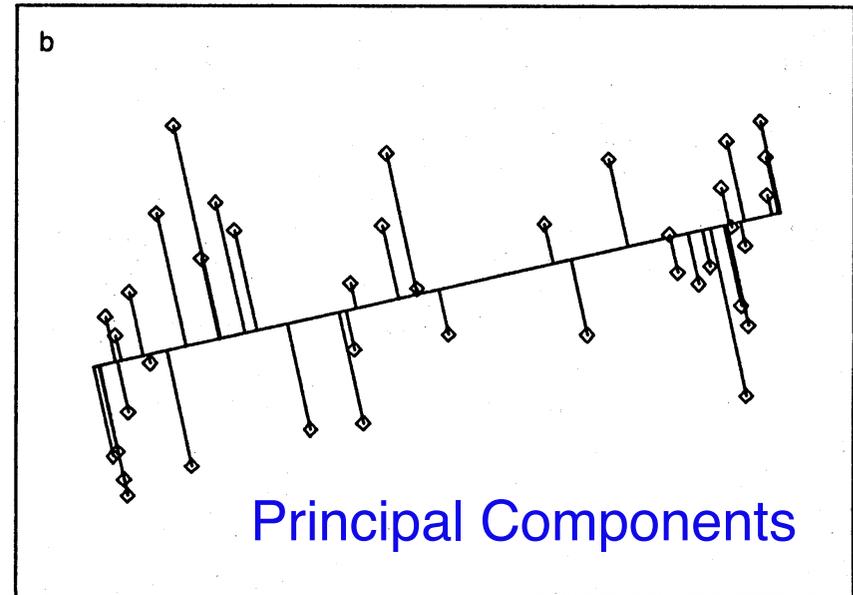
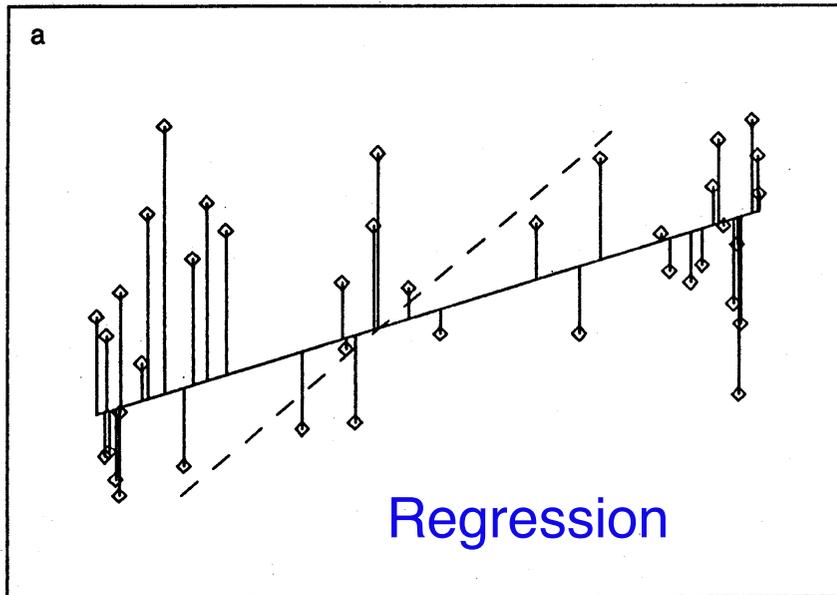
- Penalty:  $\tau \sum_{\{i, j | D_{ij} = \infty\}} \|x_i - x_j\|^2$

- Non-interacting loci cannot be too close

# Constraints and Contiguity

- Many biological constraints can be imposed:
  - Yeast: centromere clustering, 1  $\mu$ m sphere.
  - Constraints are difficult to specify; cell-type, resolution specific; increase compute burden.
  - Malaria: adjacent 10kb loci within 91nm;  
Yeast: adjacent 10kb loci  $>$  30nm.
  - Indirect way of imposing contiguity.
- Here we directly prescribe that the solution, per chromosome, is a 1D smooth curve.

# Principal Curves



# Principal Curve Metric Scaling

Goal: 1D curve  $f$  in  $R^3$  with inner products between  $n$  points on  $f$  approximating  $C_{n \times n}$ .

$f(\lambda)$ : vector fn with 3 components;  $\lambda$  1D index.  
Genomic coordinates

Want coordinate functions to be smooth wrt  $\lambda$  so we represent each using a spline basis:

$$f_{ij}(\lambda) = \sum_{k=1}^K h_{ik}(\lambda) \theta_{kj}, \quad j = 1, 2, 3; \quad i = 1, \dots, n$$

where  $K$  is the number of knots  $\sim$  spline  $df$ .

$F = H\Theta$  where  $\Theta$  is  $K \times 3$  matrix of coefficients.

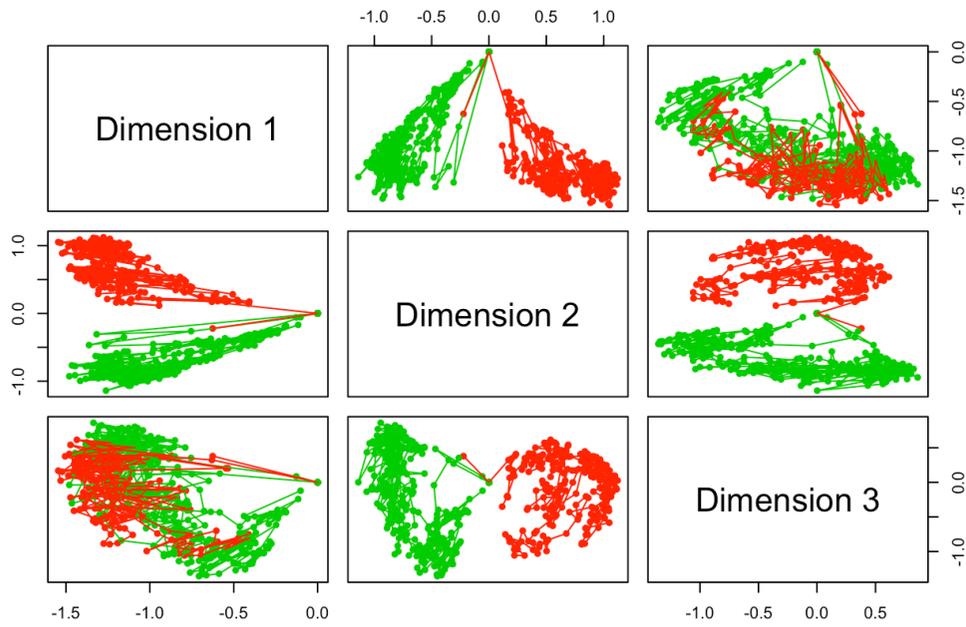
WLOG assume  $H$  is orthonormal.

Metric scaling problem:  $\min_{\Theta} \|C - H\Theta\Theta^T H^T\|_F^2$ .

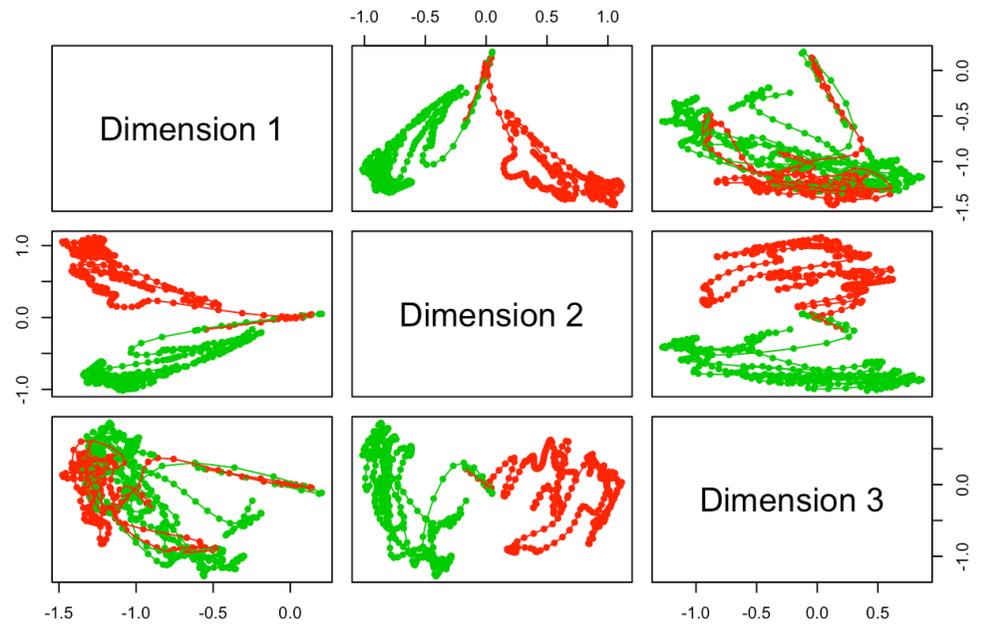
This is equivalent to  $\min_{\Theta} \|H^T C H - \Theta\Theta^T\|_F^2$

which is solved by eigen-decomposition of  $H^T C H$ .

Df = 625 R-squared = 0.78

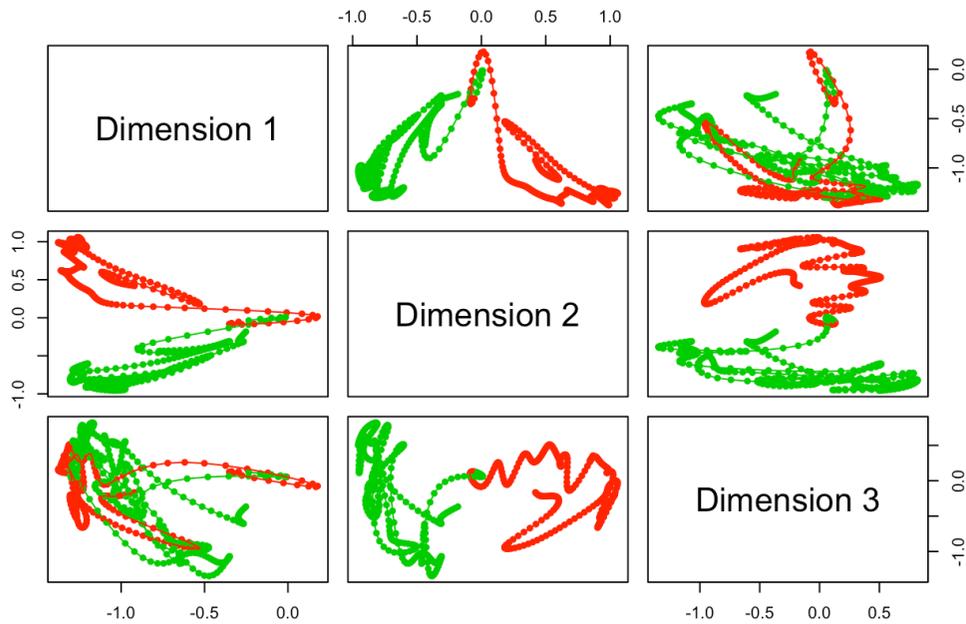


Df = 150 R-squared = 0.76

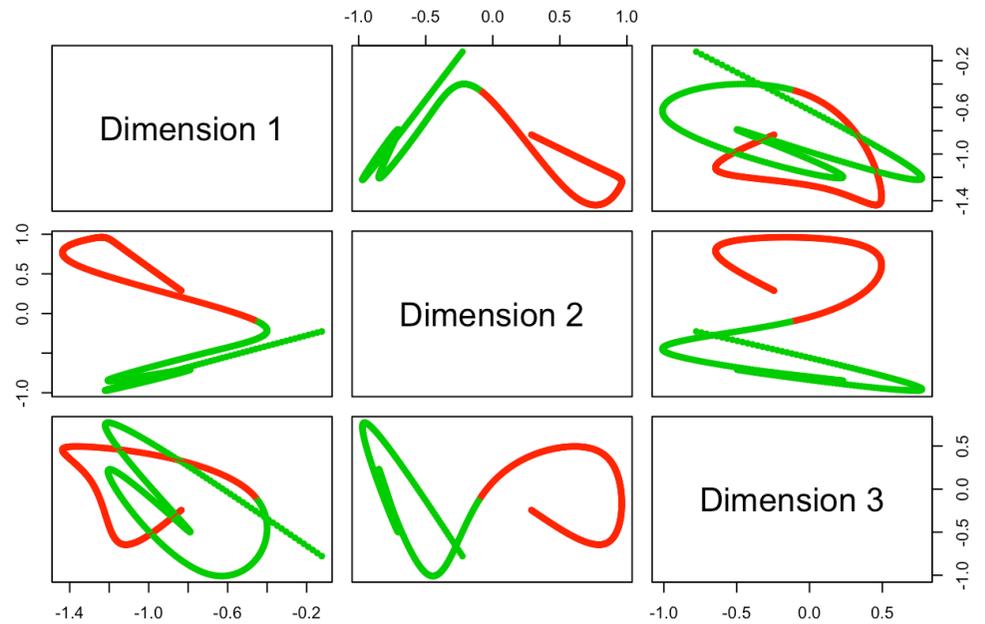


## IMR90 // Chromosome 20 // 100kb // Primary Series

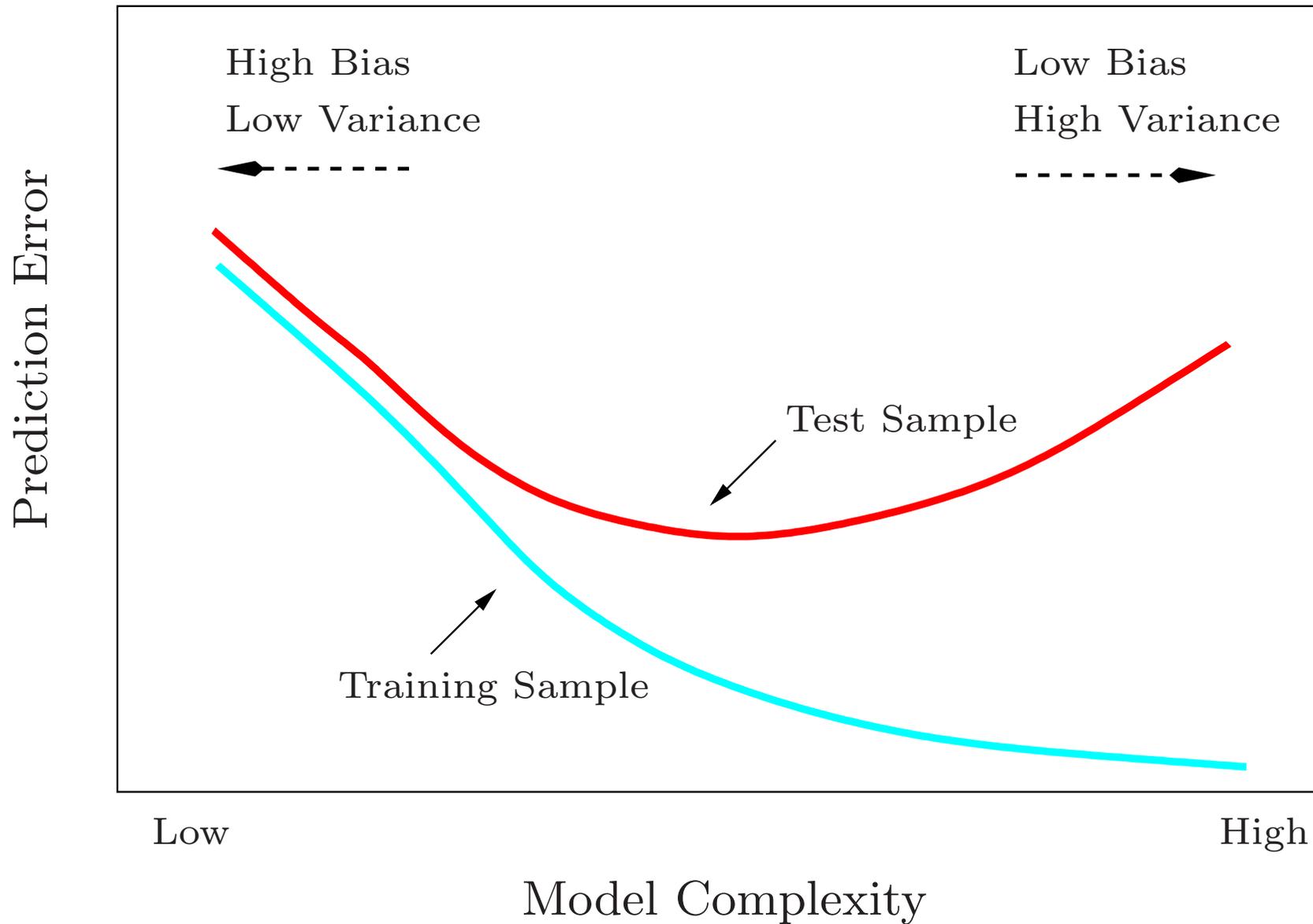
Df = 75 R-squared = 0.75



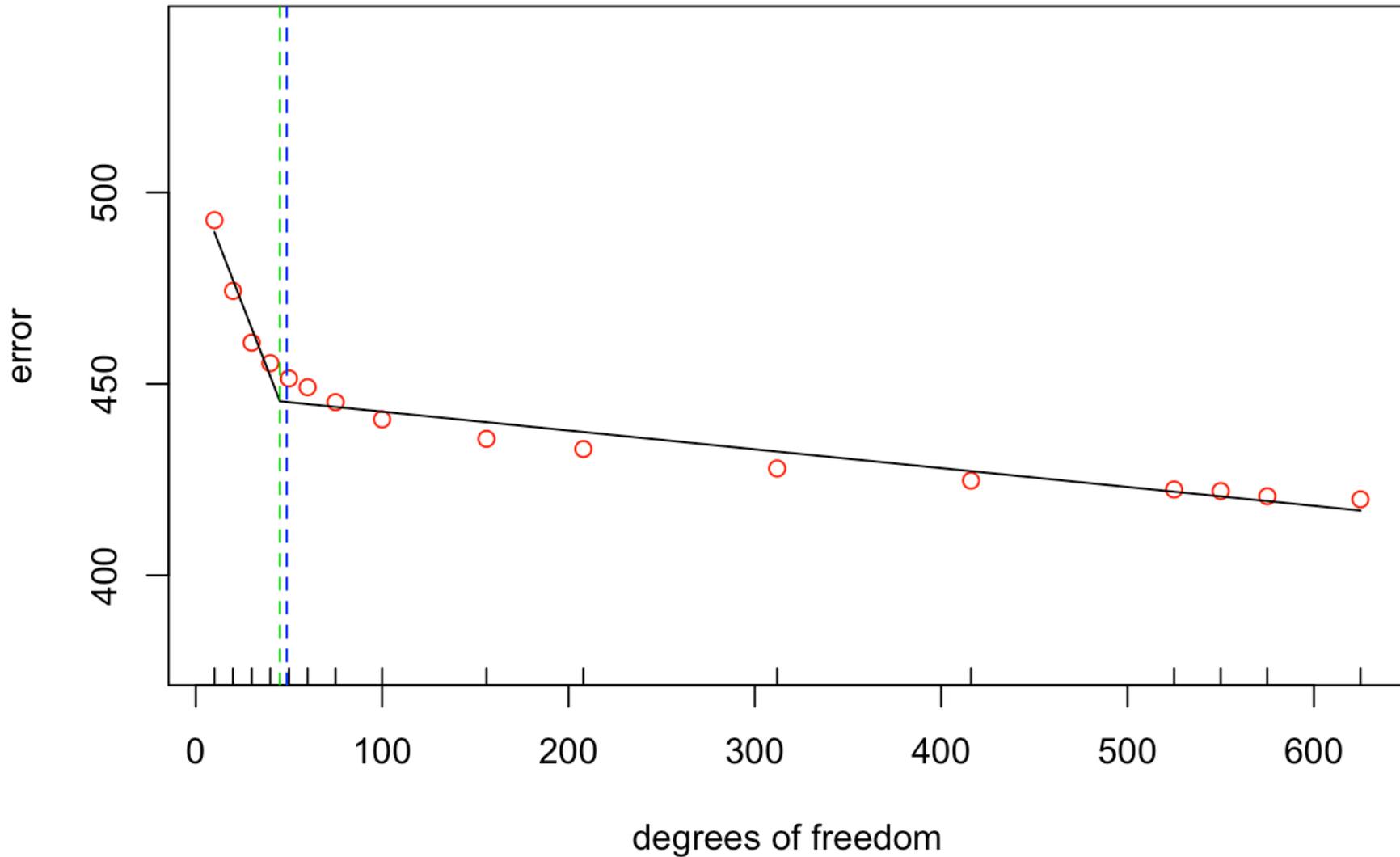
Df = 10 R-squared = 0.69



# Determining Degrees-of-Freedom



# Determining Degrees-of-Freedom

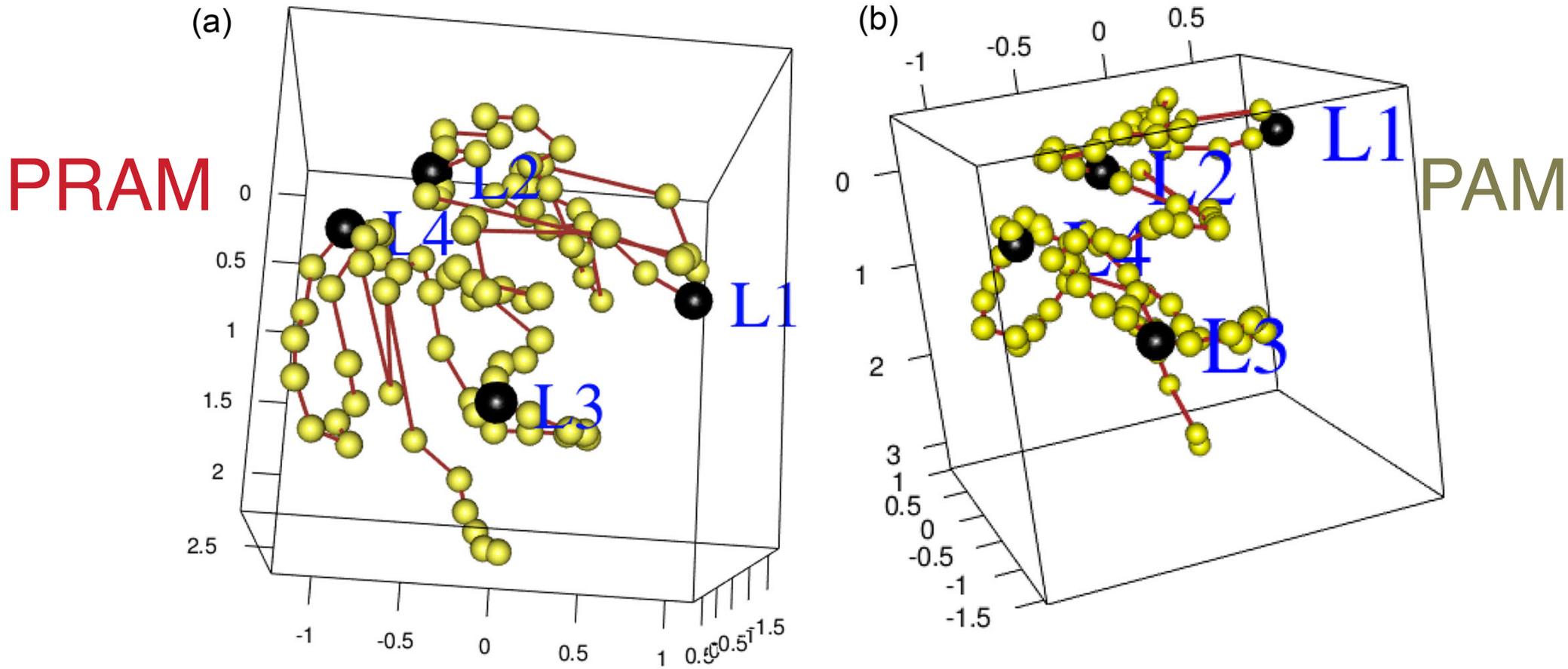


Broken-line / segmented regression: knot / elbow identification

# Assessing Reconstruction Accuracy

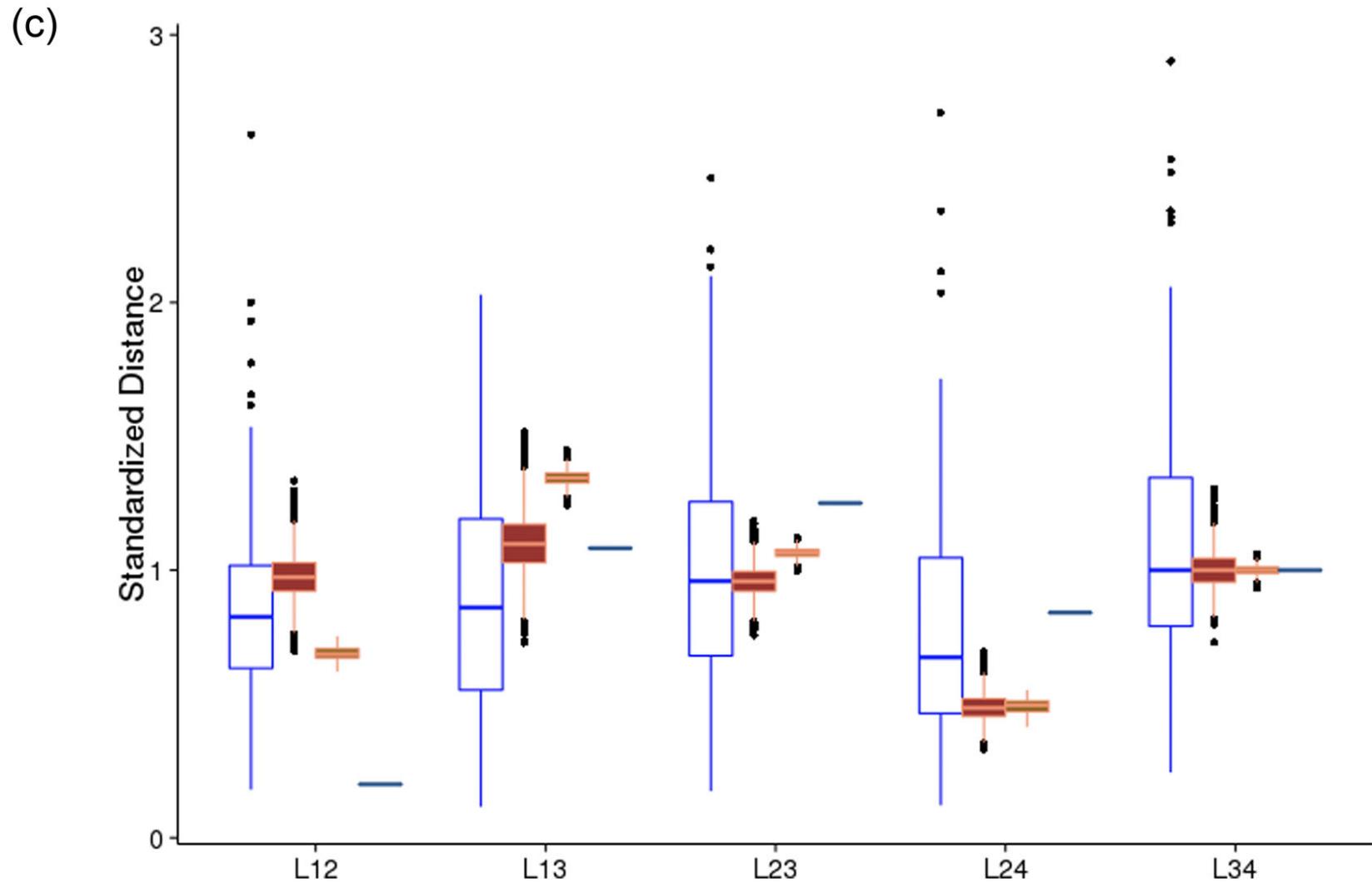
- Challenging in view of absence of gold standards
  - reproducibility assessment based on replicates from differing RE digests
- Use of FISH: compare inter-probe distances
  - exceedingly limited due to probe sparsity
- **Multiplexed FISH** affords new possibilities

# Standard FISH: 1 Mb Resolution



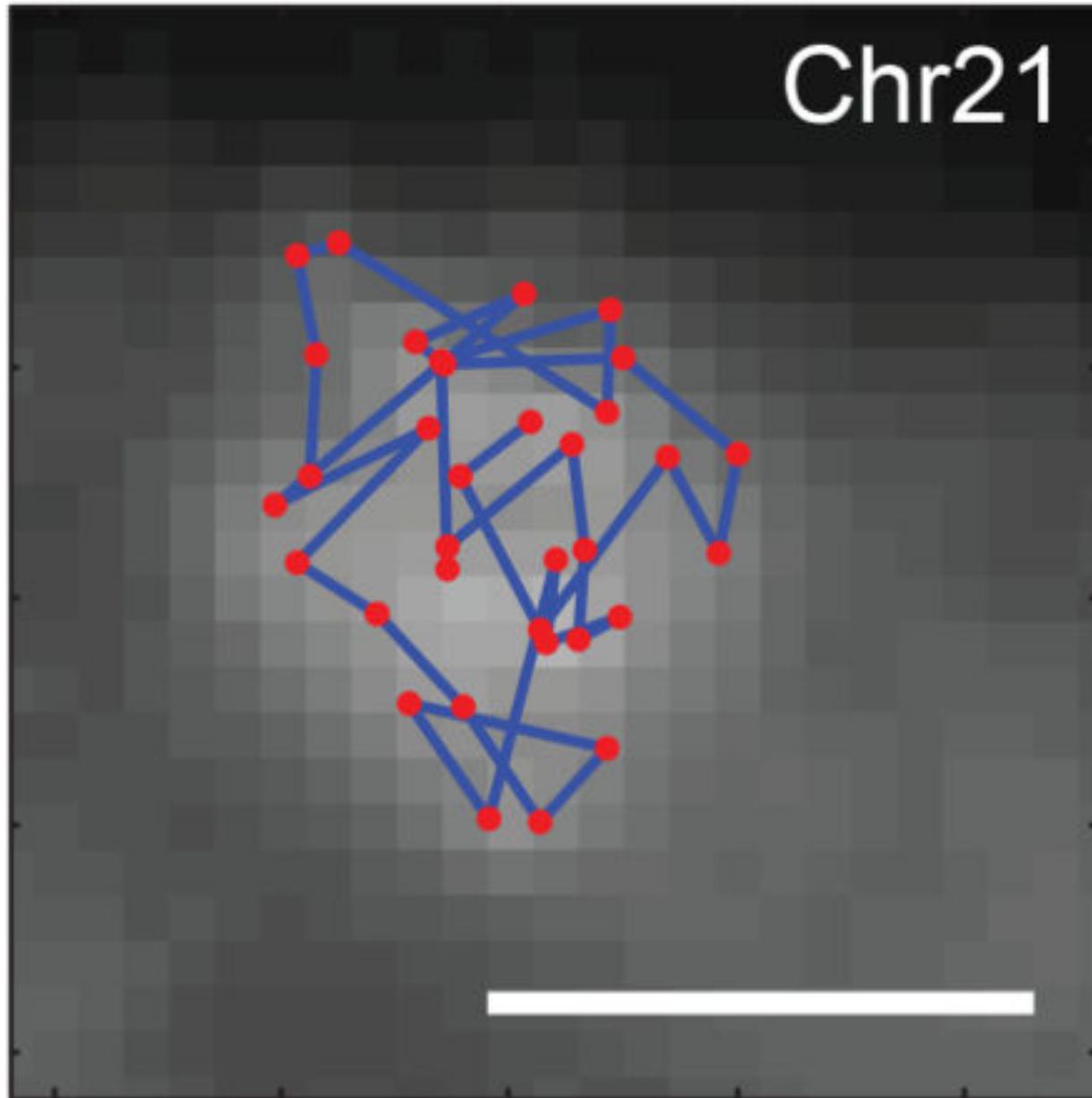
Park, Lin *Biometrics* (2016)

# Standard FISH: 1 Mb Resolution



FISH, PRAM, PAM, ShRec3D

# Multiplex FISH: 100kb Resolution



# Multiplex FISH Assessments

- Crucial is existence of numerous replicates
  - provides natural referent distribution of (R)MSD distances
  - necessary in absence of thresholds (as per protein folding) or theoretic models
- For IMR90 cells have 111, 120, and 151 replicates for chromosomes 20, 21 and 22.
- Here evaluate 3D reconstruction obtained via **PCMS** algorithm using IMR90 Hi-C data.

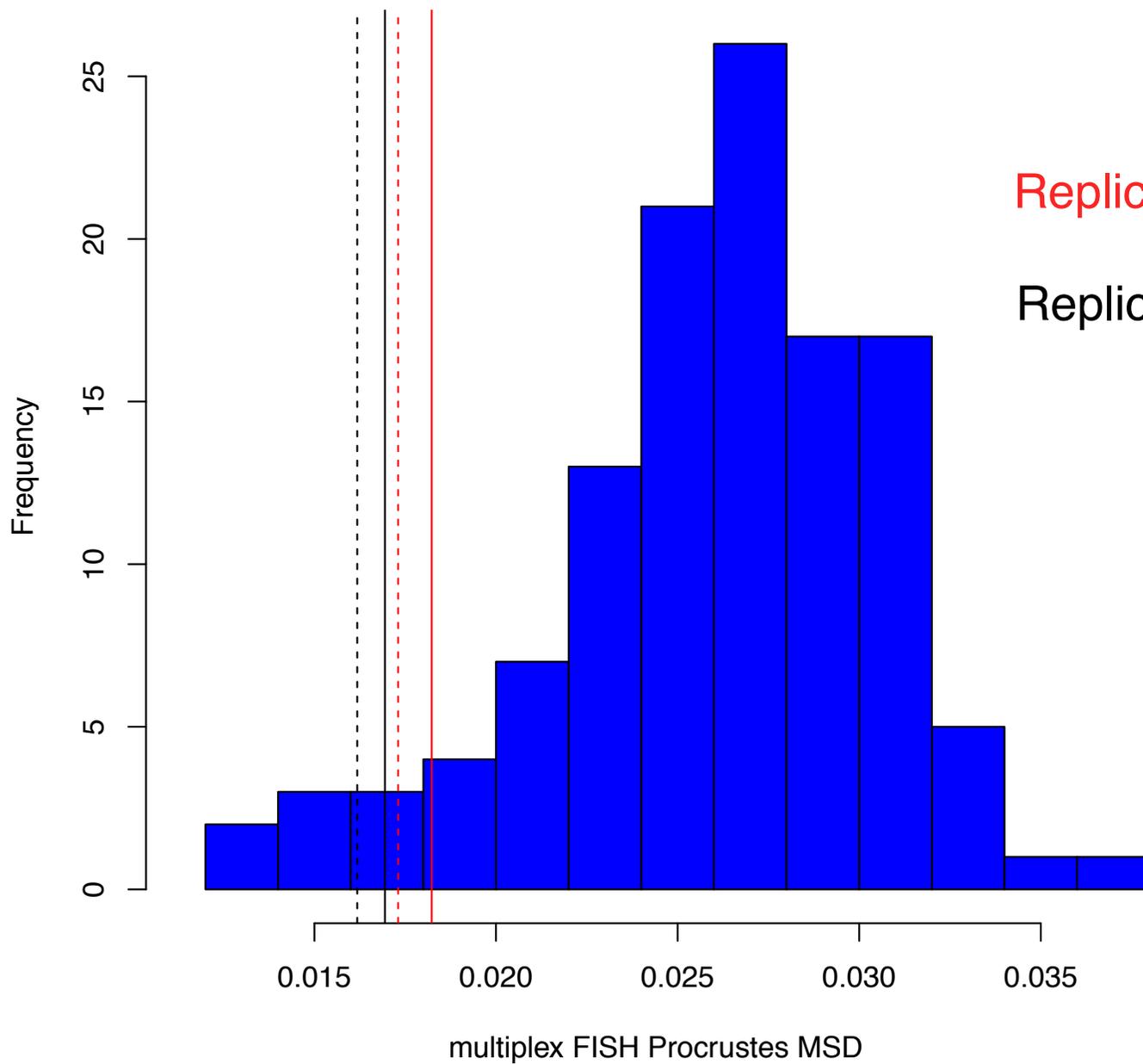
chr21 // 50kb

Primary series / Elbow  $df$

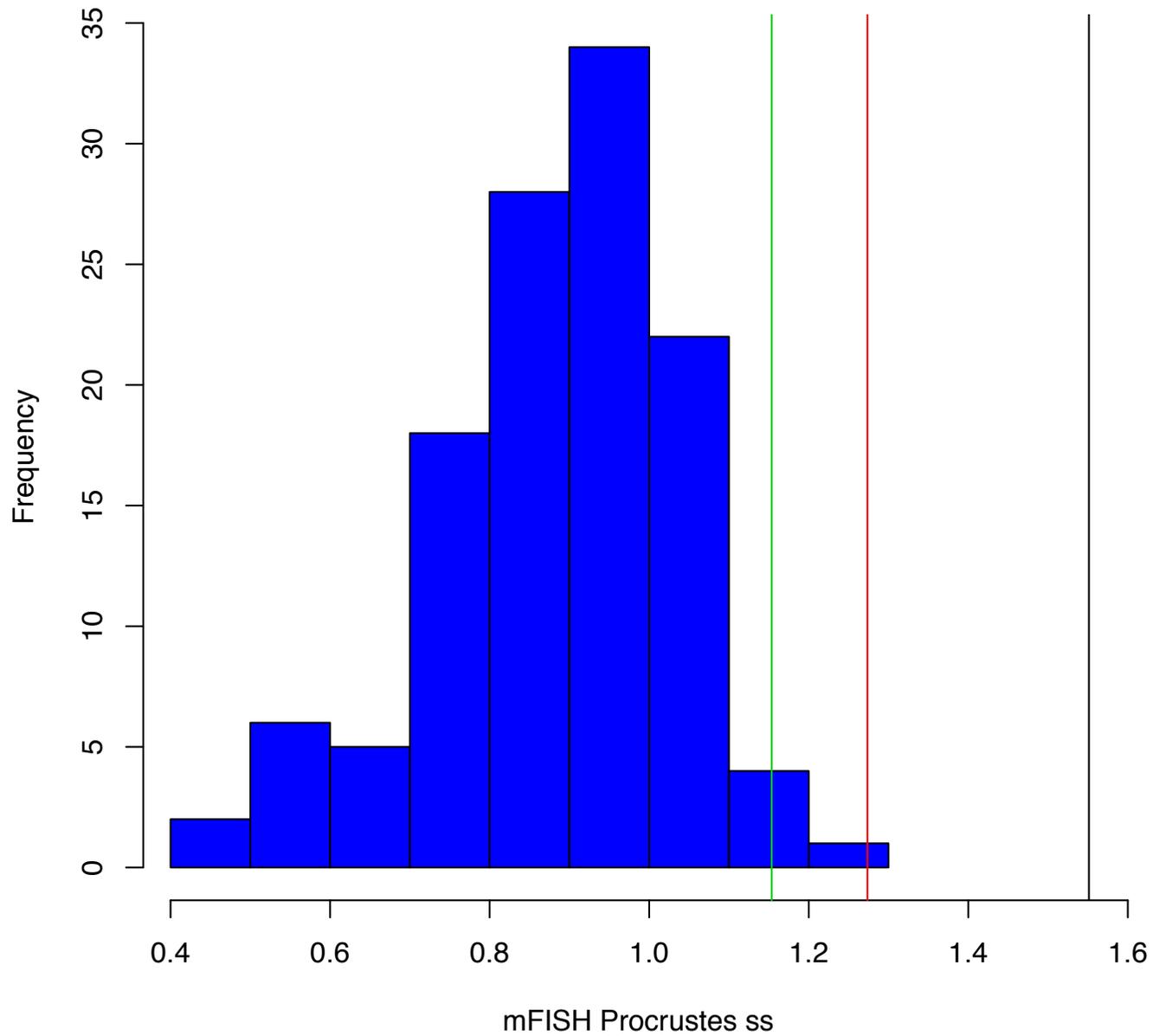
Primary series /  $df = n$

Replicate series / Elbow  $df$  - - -

Replicate series /  $df = n$  - - -



chr21 // 50kb



Alternate algorithm — HSA: **primary**, **replicate**, combined

# Future Work

- Degrees-of-freedom via cross-validation.
- Alternate bases (e.g. wavelets) or partitioning methods to capture hierarchical chromatin organization.
- Alternate transformations of  $C$ .
- Single-cell Hi-C.

# Acknowledgements

- Trevor Hastie
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