Model based clustering of longitudinal data: application to modeling disease course and gene expression trajectories

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Abstract

We consider the problem of clustering time dependent data. The model is a mixture of regressions, with variance-covariance matrices that are allowed to vary within the extended linear mixed model family. We discuss applications to biomedical data and analyze two longitudinal data sets: one on patients with delirium, and the other on mosquito gene expression following infection.

13 **1** Introduction

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¹⁴ Model based clustering (MBC) is increasingly popular as a method for studying ¹⁵ multivariate data. Most of the applications rely on the approach proposed by ¹⁶ Banfield and Raftery (1993) and implemented in the R-package MCLUST. In ¹⁷ this approach data are modeled as a mixture of multivariate normal distributions; ¹⁸ an economic parameterization of the variance-covariance matrices is achieved by ¹⁹ considering the spectral decomposition of the matrices.

New challenges appear when analyzing longitudinal data with non-negligible 20 correlations. We are interested in two broad areas of application: clustering dis-21 ease trajectories in a clinical setting, and clustering longitudinal data in gene 22 expression at several points in time. The spectral decomposition is of limited help 23 when working with such data, since it does not address the special form that the 24 variance covariance matrices may take. In addition, longitudinal data consist of 25 measurements taken repeatedly on a number of observational units, with the typ-26 ical feature, especially in clinical settings, that both the number of measurements 27 and the time points may differ across individual units. Analysis of such data is 28 usually performed using the extended linear mixed model (ELMM), see Pinheiro 29 and Bates (2000). However, the ELMM usually assumes a Gaussian distribu-30 tion for all random effects and error terms. This assumption has been relaxed 31 to include mixtures of Gaussian distributions; see, for instance, Belin and Rubin 32 (1995), Tango (1998), Trottier (1998), Verbeke and Molenberghs (2000), Luan and 33

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Li (2003), Gaffney and Smyth (2003), Celeux et al. (2005), Heard et al. (2006),
Ng et al. (2006), De la Cruz-Mesa (2008). To the best of our knowledge, however,
there is no fully developed method for simultaneously estimating parameters of
both the random effects and the error term.

While the primary methods presented in this paper have been developed else-38 where, their application is affected by several subjective choices, based on prag-39 matic considerations and on assumptions, only valid in specific contexts—these 40 are the focus of the current paper. We study a general model for representing 41 mixtures of longitudinal data that generalizes previous attempts: a mixture of 42 ELLM (Section 2). We develop an EM approach to the estimation of its param-43 eters (Section 3) and validate it through simulations (Section 4). In Section 5 is 44 we apply the model to the study of disease course, analyzing data on delirium 45 in an elderly population. Section 6 presents an application to longitudinal gene 46 expression data. Section 7 concludes the paper with a brief discussion. 47

$_{48}$ 2 The model

Let $Y_i(t_{ij})$ be the observation of the *i*th individual at time t_{ij} , for $i = 1, ..., n, j = 1, ..., m_i$, where *n* is the number of individuals and m_i is the number of time points at which the *i*th individual has been observed. The ELMM can be written as follows:

$$Y_i = X_i\beta + Z_ib_i + \epsilon_i \tag{1}$$

where X_i and Z_i are design matrices:

$$X_{i} = \begin{pmatrix} g_{1}(t_{i1}) & \dots & g_{p}(t_{i1}) \\ \dots & \dots & \dots \\ g_{1}(t_{im_{i}}) & \dots & g_{p}(t_{im_{i}}) \end{pmatrix}, \qquad Z_{i} = \begin{pmatrix} h_{1}(t_{i1}) & \dots & h_{q}(t_{i1}) \\ \dots & \dots & \dots \\ h_{1}(t_{im_{i}}) & \dots & h_{q}(t_{im_{i}}) \end{pmatrix},$$

and:

$$\beta = (\beta_1, ..., \beta_p)', \qquad b_i = (b_{i1}, ..., b_{iq})', \qquad \epsilon_i \sim N(0, \sigma^2 \Lambda_i)$$

with b_i and ϵ_i assumed independent. Here, Y_i is independent of Y_j for $i \neq j$ and Λ_i is an $m_i \times m_i$ matrix that may depend on *i* through the time intervals t_{ij} , $j=1,...,m_i$ but not otherwise. Typically, Λ_i is parameterized in terms of a relatively small number of variance parameters. Furthermore, the distribution of the random effects, b_i , is assumed to be $N(0, \Psi)$ where Ψ is a symmetric positive definite matrix which may depend on parameters to be estimated. Finally, the g_i 's and the h_i 's denote the elements of a basis in function space. In practice the columns of Z_i are often chosen as a subset of the columns of X_i . We have:

$$Y_i \mid b_i \sim N(X_i\beta + Z_ib_i, \sigma^2\Lambda_i)$$

and:

$$Y_i \sim N(X_i\beta, \Sigma_i), \quad \Sigma_i = (Z_i\Psi Z_i^T + \sigma^2\Lambda_i).$$

The random effects b_i may be considered as missing data, and maximum likelihood estimation is done by the EM algorithm (see Lindstrom and Bates (1988)). Since the joint likelihood of $(y_i^T, b_i^T)^T$ is equal to that of $((y_i \mid b_i)^T, b_i^T)^T$, we have:

$$\begin{bmatrix} y_i \mid b_i \\ b_i \end{bmatrix} \sim N\left(\begin{pmatrix} X_i\beta + Z_ib_i \\ 0 \end{pmatrix}, \begin{bmatrix} \sigma^2\Lambda_i & 0 \\ 0 & \Psi \end{bmatrix} \right)$$
(2)

Then, as is done for missing data, we can write the "complete data" log-likelihood (Trottier 1998):

$$l(\beta, \sigma^{2}, \Psi, \Lambda \mid y, b) = -\frac{1}{2} \sum_{i=1}^{n} \left(m_{i} \log(2\pi) + m_{i} \log(\sigma^{2}) + \log(|(\Psi)|) + \log(|(\Lambda_{i})|) + b_{i}^{T}(\Psi^{-1})b_{i} + \frac{(Y_{i} - X_{i}\beta - Z_{i}b_{i})^{T}(\Lambda_{i})^{-1}(Y_{i} - X_{i}\beta - Z_{i}b_{i})}{\sigma^{2}} \right).$$
(3)

Under the assumption that the n individuals are sampled from K distinct component distributions, we can write

$$Y_i = \sum_{k=1}^{K} \alpha_k (X_i \beta_k + Z_i b_{i(k)} + \epsilon_{i(k)})$$
(4)

49 where the α_k 's are the mixing coefficients:

Under this model formulation, each component, k, is distinct, uniquely defined by β_k , Ψ_k , σ_k^2 , and Λ_k . The log-likelihood of the mixture model can be defined as:

$$\sum_{i=1}^{n} \log\left(\sum_{k=1}^{K} \alpha_k \exp\left\{l_k(\beta_k, \sigma_k^2, \Psi_k, \Lambda_k \mid y_i, b_{i(k)})\right\}\right)$$
(5)

where, $l_k(\cdot|\cdot)$ is given in Equation (3). Direct maximization of the log-likelihood can be quite difficult due to the sum of terms inside the logarithm. However, we can again complete the data by considering the unobserved latent indicator variables $\delta_{i(k)}$, which is equal to 1 if observation *i* belongs to cluster *k* and 0 otherwise, and write the complete data log-likelihood (Celeux et al., 2005):

$$l = \sum_{i=1}^{n} \sum_{k=1}^{K} \left\{ \delta_{i(k)} \log(\alpha_k) + \delta_{i(k)} l_k(\beta_k, \sigma_k^2, \Psi_k, \Lambda_k \mid y_i, b_{i(k)}) \right\}$$
(6)

where l_k is as in equation (3). Thus, with this "double completion" of the data, maximum likelihood estimates of the parameter vector $\theta = (\alpha, \beta, \sigma^2, \Psi, \Lambda)$ can be obtained using an EM approach as described in the next section.

⁵³ 3 Model estimation and inference

54 3.1 EM algorithm

The EM algorithm consists of iterating until convergence between the following Eand M-steps. At iteration q > 0, the E-step consists of computing the expectation of the 'complete' log-likelihood knowing the observed data and a current value for the parameters $\theta^{[q]} = (\alpha^{[q]}, \beta^{[q]}, \Psi^{[q]}, (\sigma^2)^{[q]}, \Lambda^{[q]})$, i.e.:

$$Q(\theta \mid \theta^{[q]}) = E\left[l(\theta \mid y, \delta, b) \mid y, \theta^{[q]}\right] = \sum_{i=1}^{n} \sum_{k=1}^{K} \left(\tau_{i(k)}^{[q]} \log(\alpha_k) + \tau_{i(k)}^{[q]} E\left[l_k(\beta_k, \sigma_k^2, \Psi_k, \Lambda_k \mid y_i, b_{i(k)}) \mid y, \theta^{[q]}\right]\right)$$

where the complete data log-likelihood l is given in Equation (6), and $\tau_{i(k)}^{[q]} = E[\delta_{i(k)}|y, \theta^{[q]}]$ are the so-called posterior probabilities of component membership, computed by:

$$\tau_{i(k)}^{[q]} = P(i \in C_k \mid y_i, \theta^{[q]}, \alpha^{[q]}) = \frac{\alpha_k^{[q]} g_{m_i}(y_i \mid \theta_k^{[q]})}{\sum_{l=1}^K \alpha_l^{[q]} g_{m_i}(y_i \mid \theta_l^{[q]})}$$

where C_k denotes the k-th cluster and $g_{m_i}(y_i|\theta_k^{[q]}) = \exp(l_k(\theta_k^{[q]}|y_i))$ denotes the density of the k-th mixture component. The M-step consists of setting $\theta^{[q+1]} =$ arg max_{θ} $Q(\theta \mid \theta^{[q]})$. Details are given in Appendix A. Suffice it to say here that the M-step uses the same numerical methods for the estimation of the parameters of the Λ_i matrix as in the R functions lme and gls; therefore, though our approach follows essentially Celeux et al. (2005), it also borrows from Pinheiro and Bates (2000).

62 3.2 Initial values

It is well known that the EM algorithm can be quite sensitive to the choice of 63 starting values. A number of different strategies for choosing starting values have 64 been proposed (McLachlan and Peel, 2000). Following Celeux et al. (2005), we per-65 form a large number of short runs (10 iterations) of the EM from different k-means 66 results. The starting values which initialized the best "short run" solution (i.e. 67 the short-run solution to achieve the highest log-likelihood), are then selected as 68 starting values. When the number of observations is not equal across individuals, 69 k-means is performed on regression parameters obtained from linear regressions 70 on each individual. 71

72 3.3 Standard Errors

The asymptotic covariance matrix of the maximum-likelihood estimates, $\hat{\theta}$, is equal to the inverse of the expected Information matrix, $\mathcal{I}(\theta)$, which can be approximated by following Louis(1982)'s decomposition of $\mathcal{I}(\hat{\theta})$:

$$\mathcal{I}(\hat{\theta}) = \mathcal{E}_{\eta}(B(y,\theta)) - \mathcal{E}_{\eta}(S(y,\theta)S^{T}(y,\theta)) + \mathcal{E}_{\eta}(S(y,\theta))\mathcal{E}_{\eta}(S^{T}(y,\theta))$$
(7)

where η represents the missing data, y, the observed data and:

$$B(y,\theta) = \frac{\partial^2 \log l(\theta \mid y)}{\partial \theta^2}, \quad S(y,\theta) = \frac{\partial \log l(\theta \mid y)}{\partial \theta}$$
(8)



Figure 1: Histograms of number of clusters retrieved by the algorithm when 1 cluster is simulated using AIC(left panel) and BIC(right panel).

The standard errors of $\hat{\theta}$ are then given by the diagonal elements of $(\mathcal{I}^{-1}(\hat{\theta}))^{1/2}$. The See Appendix B for details.

78 3.4 Assessing the Number of Clusters

Assessing the "correct" number of components or clusters in finite mixture models
is a fundamental and challenging question. The minimum AIC (Akaike Information Criterion) and BIC (Bayesian Information Criterion) rules are popular choices
and both are presented in our analysis. The performance of the minimum AIC
and BIC rules is investigated in simulation studies presented in the next Section.

⁸⁴ 4 Evaluation through simulation

We performed a limited simulation study. We varied K from 1 to 7. For every 85 fixed K, we generated 100 samples from K multivariate normal distributions with 86 exchangeable variance-covariance matrices and expectations linear in time; we then 87 applied our method to estimate the parameters and chose the number of clusters 88 using both the AIC and the BIC. This was repeated 100 times. The detailed 89 forms of the distributions were chosen so as to mimic the results of the example 90 described below. We give in Figures 1, 2 and 3, the results for K = 1, 3 and 6 91 in the form of frequencies of number of retrieved class within the 100 repetitions. 92 As it can be seen, both criteria perform reasonably well, with a tendency towards 93 more conservative choices for the BIC and more liberal ones for the AIC. Though 94 we have not carried out systematic explorations beyond those reported here, our 95 experience suggests that the behaviour of the AIC and BIC is essentially the same 96 in many situations. 97



Figure 2: Histograms of number of clusters retrieved by the algorithm when 3 clusters are simulated using AIC(left panel) and BIC(right panel).



Figure 3: Histograms of number of clusters retrieved by the algorithm when 6 clusters are simulated using AIC(left panel) and BIC(right panel).

³⁸ 5 Disease Trajectory Data: Delirium

Delirium is a condition often encountered in hospitalized elderly populations. The 99 Delirium Index (DI), is a validated measure of delirium severity developed at St. 100 Mary's Hospital, (McCusker et al. 2004). DI scores range from 0 to 21 and 101 higher scores indicated more severe delirium. We used data from 229 St. Mary's 102 patients hospitalized between 1996 and 1999. Patients were evaluated with the 103 DI at enrolment, and several times during the following 15 days. Measurement 104 times were unequally spaced and differed across individuals. In order to account 105 for correlation among repeated measurements, four different models were fit: 106

Independence

$$y_i = \beta_0 + \beta_1 x_i + \epsilon_i, \qquad \epsilon_i \sim N(0, \sigma^2 Id)$$

Component	α	β	σ^2	Ψ	ϕ
1 – Steady Course	0.153	11.692, -0.015	3.912	3.428	0.438
2-Fluctuating	0.224	9.956, 0.0078	23.171	2.059	0.387
3 - Worsening	0.204	5.742, 0.032	1.340	2.602	0.001
4 - Recovery	0.139	4.451, -0.343	2.912	0.550	0.615
5 – Fluctuating Recovery	0.279	8.569, -0.316	6.105	2.750	0.215

Table 1: Parameter Estimates for 5 cluster AR(1) and random intercept solution. Except for the slopes of components 1 and 2, all parameters are significantly different from zero at the 0.05 level according to likelihood ratio tests.

Random Intercept

$$y_i = \beta_0 + \beta_1 x_i + b_i + \epsilon_i, \qquad \epsilon_i \sim N(0, \sigma^2 Id)$$

Autoregressive

$$y_i = \beta_0 + \beta_1 x_i + \epsilon_i, \qquad \epsilon_i \sim N(0, \sigma^2 R_i), \qquad R_i = AR(1)(\phi)$$

Random intercept and autoregressive

$$y_i = \beta_0 + \beta_1 x_i + b_i + \epsilon_i, \qquad \epsilon_i \sim N(0, \sigma^2 R_i), \qquad R_i = AR(1)(\phi)$$

The DI curves have a great variety of shapes and by fitting a mixture model to 107 the longitudinal data, it is our goal to reduce these shapes to a few "typical" ones 108 which may be interpreted as distinct courses of the illness. Figure 4 shows AIC 109 and BIC values for the 28 different models fit. A 5-component model with both 110 AR(1) correlation and random intercept is selected for further investigation as it 111 provides good clinical interpretation and is the best model according to the AIC. 112 The mixture model has log-likelihood of -3118.480, AIC = 6284.961 and BIC = 113 6407.968. Parameter estimates appear in Table 1. 114

115 5.1 Interpretation

The steady course component represents a course that is quite stable: the slope 116 is negligible (and non-significant at the 0.05 level) and the variance parameter 117 reasonably small. The **fluctuating course** component is similar to the first, but 118 the variance is large, suggesting that patients fluctuate around a stable state. The 119 component named **worsening** has a positive slope, indicating a DI that increases 120 in time, hence a worsening of the delirium. The remaining two components have 121 negative slopes and are therefore named **recovery**: however, in one of them we 122 have a high variance and therefore we qualify the recovery as fluctuating. From a 123 general point of view, differences in the variance parameters may seem uninterest-124 ing. However in delirium studies it is very important to identify components with 125 large fluctuations since fluctuating severity is considered a fundamental character-126 istics of 'true' delirium. 127



Figure 4: AIC and BIC for different models across K

128 5.2 Remarks

A few comments are in order. Firstly, the four models considered in this analysis do not exhaust the possibilities of the ELMM. For example we could have fitted a model with random intercept and slope; unfortunately further exploration was limited by computational power. However, we have chosen the four models that are most currently used in biostatistical practice when analyzing longitudinal data: they reflect simple and intuitive hypotheses as to how correlation might arise.

Secondly, it should be noted that, as the AIC and BIC curves show, the selection of the number of clusters depends on the model. While this may be seen as
a limitation, it is by no means an uncommon occurrence: for example this dependence.

dence is commonly observed when working with mixtures of multivariate normal
distributions and allowing hypotheses other than homoscedasticity, e.g. using the
mclust R package (Banfield and Raftery, 1993).

Thirdly, the determination of the number of classes remains a fairly subjective exercise. Indeed neither the AIC nor the BIC provide absolutely objective criteria, as is demonstrated by the numerous alternatives proposed in the litterature. In this work we have not attempted to develop new approaches, but have limited ourselves to the most popular ones.

Fourthly, though we have shown by limited simulations that the BIC works bet-146 ter than the AIC in our context, we have actually preferred to retain the 5-cluster 147 solution corresponding to the minimum AIC rather than the 2-cluster solution 148 which minimizes the BIC. This illustrates both the limits and the advantages of 149 using a certain degree of subjectivity. Indeed, the BIC of the 5-cluster solution 150 is not very different from the minimum BIC. On the other hand, a five cluster 151 solution was proposed in a previous work on delirium by Sylvestre et al. (2006), 152 who applied an exploratory approach combining principal component analysis with 153 k-means clustering. The interpretation of our 5-clusters is very similar to the in-154 terpretation of the five clusters found by these authors, yet it has the advantage 155 of being model based. 156

Finally, though we have not studied robustness systematically, we have found that the point estimates of the fixed effect coefficients are fairly stable regardless of whether or not we include the random effect and/or the AR(1) term. This is encouraging, but further explorations are desirable.

¹⁶¹ 6 Time-course gene expression data

Microarray analysis is a valuable tool in molecular biology, as it permits to assess 162 the expression levels of a large number of genes simultaneously. In view of the 163 complexity of biological networks, it is useful to study gene expression not only 164 at a specific point in time, as in early microarray experiments, but also longitu-165 dinally. Expression time profiles can indeed be very useful to find co-regulated 166 and functionally related groups of genes. We analysed a set of longitudinal gene 167 expression data already studied by Heard et al. (2002). The data consists of 2771 168 gene expression time profiles (each with 6 non-equally spaced observations) from 169 mosquitoes which have been infected with a bacterial agent. Visualization of the 170 raw data is not very informative (Figure 5). 171

Heard et al. (2002) proposed a Bayesian model-based hierarchical clustering 172 algorithm to cluster genes having similar expression profiles that led to a 17-cluster 173 solution. From this, interpretable graphs were obtained. Their solution assumed 174 data to be uncorrelated, so that in practice their model is a mixture of ordinary 175 regressions. In contrast, we used the wealth of submodels within the ELMM to find 176 a non-trivial correlation structure that fits the data. To model the trajectories, we 177 used a flexible family of basis functions called the truncated power spline basis, as 178 in Heard et al. (2002): 179



Figure 5: and heat map of the *Salmonella typhi* data presented in Heard et al. (2002)

$$g_1(t_{is}) = 1$$
 $g_j(t_{is}) = (t_{is} - t_{i,j-1})_+, s = 1, ..., n, j = 2, ..., m_i$

where $(\cdot)_+$ is the positive part function.

We fit three different correlation structures: an independence model, an au-181 toregressive model as well as a model with random intercept and autoregressive 182 structure. We varied K from 1 to 26. According to the BIC, see Figure 6, the 183 best model is a 14-cluster model with both random intercept and autoregressive 184 structure. A heatmap of the data is presented in Figure 7. Comparing the fitted 185 values(right) across clusters gives a visual measure of between cluster heterogene-186 ity. Comparing the fitted values(right) to the raw observations(left) gives a sense 187 of within cluster homogeneity. The clustering is shown in more detail in Figure 8. 188

189 7 Discussion

We have presented a straightforward method for modeling heterogeneity in longi-190 tudinal data. We have proposed a mixture of regressions with components in the 191 Extended Linear Mixed Model (ELMM) (Pinheiro and Bates, 2000). The ELMM 192 consists of a random effect portion (LMM) extended by the addition of an error 193 term with correlation matrix defined up to a small number of parameters to be 194 estimated from data. We have limited ourselves to an autoregressive error term. 195 Our approach to parameter estimation is based on the EM algorithm of Celeux 196 et al. (2005) for mixtures of LMM, augmented by numerical methods which are 197 essentially those used in the lme and gls R functions of Pinheiro and Bates (2000). 198 The theoretical novelty of this approach is modest: it permits, on the one hand, 199 to deal with correlated errors of a type that is important in applied research, and, 200 on the other, suggests further extension to a catalog of possible error correlation 201 structures such as those contained in the lme and gls R functions. Although other 202 authors have considered mixtures of regressions for longitudinal data, no one has 203



Figure 6: log-likelihood, AIC and BIC plots for different models(independence(red), autoregressive(green) and autoregressive with random intercept(blue)) fit for K = 1,...26



Figure 7: Clustered gene expression profiles form the Salmonella typhi data

yet achieved the generality that can be achieved with the ELMM. An appropriate modeling of the correlation structure of longitudinal data is important: not only does it provide useful insight into the dynamical process under study, but it also leads to fewer clusters, hence to a more economical model of the data. We



Figure 8: 14 cluster solution suggested by autoregressive AR(1) model with random intercept model. BIC = 14528.19

have also shown that mixtures of regressions offer an important tool in classifying course of diseases from clinical data and longitudinal gene expression data,
providing easy-to-interpret analyses.

Further research will aim to speed up the EM algorithm, which will also allow us to study even richer correlation structures. Amelioration of computing efficiency will allow us to carry out more extensive simulations and to study the robustness of key features of our models, e.g. fixed effect parameter estimates and selection of the number of clusters. We plan also to revisit the Bayesian approach of Heard et al. (2002).

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²⁵⁷ A Details on the M-step of the EM algorithm

We have for the q+1 th step:

$$\alpha_k^{[q+1]} = \sum_{i=1}^n \frac{\tau_{i(k)}^{[q]}}{n}$$

$$\beta_{k}^{[q+1]} = \left[\left(\sum_{i=1}^{n} \tau_{i(k)}^{[q]} \left((y_{i} - Z_{i} b_{i(k)}^{[q]}) \right)^{T} \left((\Lambda_{ik}^{[q]})^{-1} \right) (-X_{i}) \right) \left(\sum_{i=1}^{n} \tau_{i(k)}^{[q]} \left(X_{i} \right)^{T} \left((\Lambda_{ik}^{[q]})^{-1} \right) (-X_{i}) \right)^{-1} \right]^{T} \right]^{T} \left((\sigma_{k}^{2})^{[q+1]} = \frac{1}{\sum_{i=1}^{n} \tau_{i(k)^{[q]}} m_{i}} \sum_{i=1}^{n} \tau_{i(k)^{[q]}} \left[E(e_{ik}^{T} \Lambda_{ik}^{-1} e_{ik} \mid y_{i}, \theta^{[q-1]}) \right]$$

where

$$E(e_{ik}^{T}\Lambda_{ik}^{-1}e_{ik} \mid y_{i}, \theta^{[q-1]}) = \sigma_{k}^{4[q-1]}(y_{i} - X_{i}\beta_{k}^{[q-1]})^{T}\Sigma_{ik}^{-1[q-1]}\Lambda_{ik}^{[q-1]}\Sigma_{ik}^{-1[q-1]}(y_{i} - X_{i}\beta_{k}^{[q-1]}) + m_{i}\sigma_{k}^{2[q-1]} - \sigma_{k}^{4[q-1]}tr(\Sigma_{ik}^{-1[q-1]}\Lambda_{ik}^{[q-1]})$$

and

$$\Psi_k^{[q+1]} = \frac{1}{\sum_{i=1}^n \tau_{i(k)}^{[q]} m_i} \sum_{i=1}^n \tau_{i(k)}^{[q]} E(b_{i(k)} b_{i(k)}^T \mid y_i, \theta^{[q-1]})$$

where

$$E(b_{i(k)}b_{i(k)}^{T} \mid y_{i}, \theta^{[q-1]}) = E(b_{i(k)} \mid y_{i}, \theta^{[q-1]})E(b_{i(k)} \mid y_{i}, \theta^{[q-1]})^{T} + m_{i}\Psi_{k}^{[q-1]} - \Psi_{k}^{[q-1]}Z_{i}^{T}\Sigma_{ik}^{-1[q-1]}Z_{i}\Psi_{k}^{[q-1]}$$

and

$$E(b_{i(k)} \mid y_i, \theta^{[q-1]}) = \Psi_k^{[q-1]} Z_i^T \Sigma_{ik}^{-1[q-1]} (y_i - X_i \beta_k^{[q-1]})$$

Finally, consider the positive-definite matrices Λ_{ik} (there are nK of these). There are different ways that such matrices may be parametrized, depending on assumptions regarding the intra-individual covariance structure (Pinheiro and Bates, 2000). Let ϕ_k denote the set of parameters used in the parametrization of $\{\Lambda_{ik}\}_{i=1,...,n}$. To estimate these parameters at iteration [q], we use numerical maximization methods (e.g the R function nlminb()).

²⁶⁴ B Details on computing the standard error of $\hat{\theta}$

Jennrich and Schluchter (1986) provide equations for the required score vector statistics and Hessian matrix in the homogeneous model. The required first and second derivatives for the mixture model are presented bellow. Derivatives with respect to the parameters that define Λ , (ϕ) , must be calculated by numerical methods.

$$\frac{\partial l(\theta \mid Y)}{\partial \alpha_k} = \sum_{i=1}^n \frac{\delta_{i(k)}}{\alpha_k} - \frac{\delta_{i(K)}}{\alpha_K}, \quad k=1,\dots,K-1$$
(9)

$$\frac{\partial l(\theta \mid Y)}{\partial \beta_k} = \sum_{i=1}^n \delta_{i(k)}(X_i^T)(\Sigma_{ik}^{-1}) \left(y_i - X_i \beta_k\right), \quad _{k=1,\dots,K}$$
(10)

$$\frac{\partial l(\theta \mid Y)}{\partial \sigma_k^2} = \sum_{i=1}^n \frac{\delta_{i(k)}}{2} \operatorname{tr}(\Sigma_{ik}^{-1}(y_i - X_i\beta_k)(y_i - X_i\beta_k)\Sigma_{ik}^{-1}\Lambda^{-1}), \quad _{k=1,\dots,K}$$
(11)

$$\frac{\partial l(\theta \mid Y)}{\partial \Psi_k} = \sum_{i=1}^n \frac{\delta_{i(k)}}{2} \operatorname{tr}(\Sigma_{ik}^{-1}(y_i - X_i\beta_k)(y_i - X_i\beta_k)\Sigma_{ik}^{-1}Z_iZ_i^T), \quad k=1,\dots,K$$
(12)

$$\frac{\partial^2 l(\theta \mid Y)}{\partial \alpha_k^2} = \sum_{i=1}^n \frac{-\delta_{i(k)}}{\alpha_k^2} - \frac{\delta_{i(K)}}{\alpha_K^2}, \quad k=1,\dots,K-1$$
(13)

$$\frac{\partial^2 l(\theta \mid Y)}{\partial \beta_k^2} = \sum_{i=1}^n -\delta_{i(k)} (X_i^T \Sigma_{ik}^{-1} X_i), \quad _{k=1,\dots,K}$$
(14)

$$\frac{\partial^2 l(\theta \mid Y)}{\partial (\sigma_k^2)^2} = \sum_{i=1}^n \frac{-\delta_{i(k)}}{2} \operatorname{tr}(\Sigma_{ik}^{-1} \Lambda_{ik} \Sigma_{ik}^{-1} (2(y_i - X_i \beta_k)(y_i - X_i \beta_k)^T - \Sigma_{ik}) \Sigma_{ik}^{-1} \Lambda_{ik}), \quad k=1,\dots,K$$
(15)

$$\frac{\partial^2 l(\theta \mid Y)}{\partial \sigma_k^2 \partial \beta_{jk}} = \sum_{i=1}^n -\delta_{i(k)} X_{jk} (\Sigma_{ik}^{-1} \Lambda_{ik} \Sigma_{ik}^{-1} (y_i - X_i \beta_k)), \quad k=1,\dots,K \quad j=1,\dots,p$$
(16)

$$\frac{\partial^2 l(\theta \mid Y)}{\partial \Psi_k^2} = \sum_{i=1}^n \frac{-\delta_{i(k)}}{2} \operatorname{tr}(\Sigma_{ik}^{-1} Z_i Z_i^T \Sigma_{ik}^{-1} (2(y_i - X_i \beta_k)(y_i - X_i \beta_k)^T - \Sigma_{ik}) \Sigma_{ik}^{-1} Z_i Z_i^T), \quad k = 1, \dots, K$$
(17)

$$\frac{\partial^2 l(\theta \mid Y)}{\partial \Psi_k \partial \beta_{jk}} = \sum_{i=1}^n -\delta_{i(k)} X_{jk} (\Sigma_{ik}^{-1} Z_i Z_i^T \Sigma_{ik}^{-1} (y_i - X_i \beta_k)), \quad k=1,\dots,K \quad j=1,\dots,p$$
(18)