Controlling and Testing Horizontal Pleiotropy with Probabilistic Mendelian Regression for Transcriptome-wide Association Studies

Xiang Zhou

John G. Searle Assistant Professor Department of Biostatistics University of Michigan

Transcriptome-wide Association Studies

- Genome-wide association studies (GWASs) have identified many genetic variants associated with diseases and complex traits.
- Expression quantitative trait loci (eQTL) mapping studies have also identified enabled accurate measurements of gene expression levels.
- Integrative analysis of GWASs and eQTL mapping studies has the potential to yield insight into the causal relationship between genes and complex traits.



Gene expression decomposition

PrediXcan GReX Genetically regulated expression Trait Traitaltered Other component factors

"SNP aggregation approach"

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Step 1: Construct a genetic predictor of gene expression using ElasticNet

Step 2: Test the association between genetic predictor of expression and trait





"expression-trait associations"



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Zhu et al., Nature Genetics, 2016



"identify causal genes"

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"identify causal genes"

Step 1: Construct a genetic predictor of gene expression using linear regression with one SNP

Step 2: Test the association between genetic predictor of expression and trait

- These existing approaches can all be thought of as a two-stage regression version of Mendelian randomization (MR) analysis.
- MR is a form of instrumental variable analysis with SNPs serving as instruments.
- MR is a powerful statistical tool to determine causal relationship between an exposure variable (in this case, gene expression) and an outcome variable (in this case, complex trait) in observational studies









Pervasive Horizontal Pleiotropy



Corrected: Publisher Correction

Detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization between complex traits and diseases

Marie Verbanck^{1,2,3,7}, Chia-Yen Chen^{1,2,3,6,7}, Benjamin Neale^{1,4,5,6,8*} and Ron Do^{1,2,3,8*}

• Sample I, the observed gene expression data:

$$\boldsymbol{x} = \boldsymbol{\mu}_{\mathrm{x}} + \boldsymbol{Z}_{\mathrm{x}}\boldsymbol{\beta} + \boldsymbol{\varepsilon}_{\mathrm{x}} \qquad (1)$$

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• Sample II, the unobserved gene expression data:

• Sample I, the observed gene expression data:

$$\boldsymbol{x} = \boldsymbol{\mu}_{\mathrm{x}} + \mathbf{Z}_{\mathrm{x}}\boldsymbol{\beta} + \boldsymbol{\varepsilon}_{\mathrm{x}} \qquad (1)$$

• Sample II, the unobserved gene expression data:

$$\widetilde{\boldsymbol{x}} = \boldsymbol{\mu}_{\mathrm{x}} + \boldsymbol{Z}_{\mathrm{y}}\boldsymbol{\beta} + \boldsymbol{\varepsilon}_{\mathrm{y}} \qquad (2)$$

• Sample I, the observed gene expression data:

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$$\widetilde{\boldsymbol{x}} = \boldsymbol{\mu}_{\mathrm{x}} + \boldsymbol{Z}_{\mathrm{y}}\boldsymbol{\beta} + \boldsymbol{\varepsilon}_{\mathrm{y}} \qquad (2)$$

• Sample II, the observed GWAS data:

• Sample I, the observed gene expression data:

$$\boldsymbol{x} = \boldsymbol{\mu}_{\mathrm{x}} + \mathbf{Z}_{\mathrm{x}}\boldsymbol{\beta} + \boldsymbol{\varepsilon}_{\mathrm{x}} \qquad (1)$$

• Sample II, the unobserved gene expression data:

$$\widetilde{\boldsymbol{x}} = \boldsymbol{\mu}_{\mathrm{x}} + \boldsymbol{Z}_{\mathrm{y}}\boldsymbol{\beta} + \boldsymbol{\varepsilon}_{\mathrm{y}} \qquad (2)$$

• Sample II, the observed GWAS data:

$$\mathbf{y} = \mu_{y} + \widetilde{\mathbf{x}}\alpha + \mathbf{Z}_{y}\mathbf{\gamma} + \boldsymbol{\epsilon} \qquad (3)$$

Additional Modeling Assumptions

- Because the number of SNPs (p) is often larger than the sample size (n), we need to make additional modeling assumption for model identifiability.
- For β , we follow standard polygenic models to assume $\beta_i \sim N(0, \sigma_\beta^2)$.

• For γ , we follow Egger regression to assume $\gamma_1 = \cdots = \gamma_p = \gamma$

Probabilistic Mendelian Randomization

- Instead of the usual two-stage regression procedure, we rely on the maximum likelihood estimation procedure for inference.
- We develop a computationally efficient fitting algorithm, based on a parameter expansion version of the expectation maximization algorithm (PX-EM).
- We test causal effect H_0 : $\alpha = 0$ through LRT.
- We test horizontal pleiotropic effect $H_0: \gamma = 0$ through LRT.
- We refer to our method as PMR-Egger.

Simulations

- We extracted p = 556 cis-SNPs of a gene from the GEUVADIS data $(n_1 = 465)$ and simulated gene expression.
- We extricated the same SNPs from 2,000 controls in the Wellcome trust case control consortium (WTCCC) and simulated trait.
- We examined various scenarios, with 10,000 replicates for each scenario.

Compared Methods: Testing α

- PrediXcan: Elastic Net prior on β ; no γ ; two-stage inference
- TWAS: BSLMM prior on β ; no γ ; two-stage inference
- SMR: Single β ; no γ ; two-stage inference
- CoMM: Normal prior on β ; no γ ; maximum likelihood inference
- LDA MR Egger: Fixed effects of $oldsymbol{eta}$; Egger assumption on $oldsymbol{\gamma}$; two-stage inference
- PMR-Egger: Normal prior on $\boldsymbol{\beta}$; Egger assumption on $\boldsymbol{\gamma}$; maximum likelihood inference

Testing Causal Effect α under the Null



 $\boldsymbol{\gamma}=\boldsymbol{0}$



 $\gamma = 0.001$

Violation of the Polygenic β Assumption

 $\gamma = 0.001$

 $\gamma = 0.002$



Violation of the Homogeneous γ Assumption

 $\gamma = 0.001$

 $\gamma = 0.002$



Power of Testing α under Alternative



Compared Methods: Testing γ

- LDA MR Egger: Fixed effects of β ; Egger assumption on γ ; two-stage inference.
- MR-PRESSO: Permutation based approach; assumes independent instruments.
- PMR-Egger: Normal prior on β ; Egger assumption on γ ; maximum likelihood inference.

Testing Horizontal Pleiotropy γ under the Null

 $PVE_{zy} = 0.004$

 $PVE_{zy} = 0.006$



Power of Testing γ under the Alternative



Real Data Applications

• GEUVADIS Expression Data ($n_1 = 465$), with ~15,000 genes.

• WTCCC: Seven common diseases ($n_2 = \sim 5,000$).

• UK Biobank: Ten quantitative traits ($n_2 = \sim 300,000$).

WTCCC: Testing Causal Effects



WTCCC: Testing Horizontal Pleiotropy



Expected (-log₁₀ p-value)

BD

Expected (-log₁₀ p-value)

T1D

UK Biobank: Testing Causal Effects



Expected -log10(p-value)

UK Biobank: Testing Horizontal Pleiotropy



Summary

- We have presented an MR framework that unifies many existing integrative transcriptome wide association analysis method.
- Our method PMR-Egger effectively controls for horizontal pleiotropy through a maximum likelihood/probabilistic inference framework.
- We have demonstrated the effectiveness of PMR-Egger through simulations and real data applications.
- PMR-Egger is implemented in the PMR R package, to be available on <u>www.xzlab.org</u>

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